Journal of American Science

Websites: http://www.jofamericanscience.org http://www.sciencepub.net

Emails: editor@sciencepub.net sciencepub@gmail.com



Study of the effects of some oral Anti-diabetic drugs and Silymarin on Experimentally Induced Non-alcoholic Fatty Liver in Rats

Rashad Abd El-Nabi Atlam, Hammouda Hassan Sharaf and Amin Yehia Awad

Pharmacology Department, Faculty of Medicine for Male, Al-Azhar University (Cairo), Egypt Email: rasatlam@gmail.com

Abstract: Nonalcoholic fatty liver disease (NAFLD) is a chronic disease which has a wide spectrum of liver-related histological damage and becoming a global epidemic. Predisposing factors for NAFLD and related complications include insulin resistance and type 2 diabetes mellitus. Therefore, intervention with anti-diabetic medications may prevent and delay the development of NAFLD or have a therapeutic application. The focus of this study is to evaluate the evidence supporting the efficacy of anti-diabetic medications and silymarin in the treatment of NAFLD. Methods: 48 male albino rats divided in to 4 groups: Group I received standard diet, Group II received a high fat diet (HFD), Group III treated with empagliflozin (10 mg/ kg/ day) and Group IV treated with silymarin (100 mg/ kg/ day). After 5 weeks rats were scarified and serum (ALT, AST, TG, cholesterol, random blood glucose and insulin level), hepatic (MDA, GSH, TG and TGF- β) and body weight were measured. Histopathological examination was done. Results: empagliflozin, and silymarin had good results in ameliorating NAFLD as regard anti-inflammatory, anti-hyperlipidemic, anti-fibrotic effects and decreasing hepatic lipid content. Clinically such findings may suggest that, these drugs could be used in treatment of NAFLD patients especially those have type 2 DM and liver fibrosis. [Rashad Abd El-Nabi Atlam, Hammouda Hassan Sharaf and Amin Yehia Awad. Study of the effects of some oral Anti-diabetic drugs and Silymarin on Experimentally Induced Non-alcoholic Fatty Liver in Rats. J Am Sci 2020;16(12):107-123]. ISSN 1545-1003 (print); ISSN 2375-7264 (online). http://www.jofamericanscience.org. 10. doi:10.7537/marsjas161220.10.

Keywords: NAFLD; NASH; anti-diabetic medication; silymarin.

1. Introduction

Nonalcoholic fatty liver disease (NAFLD) is the most common cause of liver disease currently worldwide. NAFLD is a major cause of chronic liver disease. Fatty liver ranges from simple triglyceride accumulation (steatosis), non-alcoholic steato-hepatitis (NASH) to the most serious, cirrhosis and also hepatocellular carcinoma. It is reported that almost 10-20% of individuals with NAFLD have NASH and 10-15% of individuals with NASH develop cirrhosis (Alam et al., 2018).

The pathogenesis of NAFLD is complex, involving insulin resistance, oxidative stress, lipid peroxidation and mitochondrial dysfunction. Insulin resistance is the most important pathogenic factor for the development of NAFLD (**Kuchay et al., 2018**).

High-fat diet is the commonest cause of obesity in the world. Diet-induced weight loss and life style modifications, including physical exercise and qualitative changes in the diet, have significant effects on NAFLD and antagonizes insulin resistance (IR). However, only a small percentage of patients with NAFLD can apply these measures efficiently (**Souza** et al., 2010). Empagliflozin is a potent oral anti-diabetic drug that inhibits sodium-glucose co-transporter 2 (SGLT-2). By inhibiting this transporter, empagliflozin promotes urinary glucose excretion, resulting in decrease in blood levels of glucose and improves insulin resistance in patients with type 2 diabetes leading to down-regulation of carbohydrate-responsive element-binding protein (ChREBP), a transcription factor responsible for activating the machinery for fatty acid synthesis. Improvement in insulin resistance (hyperinsulinemia) results in down-regulation of SREBP-1c and the blockage of hepatic lipogenesis (Kuchay et al., 2018).

Silymarin has many hepato-protective properties such as anti-inflammatory, anti-proliferative, immunemodulatory and anti-cholesterolemic characters. So, silymarin has been used in the treatment of many liver disorders (Marin et al., 2017).

2. Material and Methods:

Animal grouping and design of the work:

48 male albino rats each weighing 100-120 grams were kept under similar housing conditions and were divided in to 4 groups as the following:

• Group I (serving as negative control group):

12 rats were fed with standard diet and received normal saline at a dose of 1 ml/ rat by gastric tube daily throughout the study, and served as normal control group.

• Group II (serving as NAFLD control group):

12 rats were fed a high fat diet (HFD); which consists of the standard crushed rat chow diet, 10% animal fat (sheep tallow) and 2% cholesterol for 16 weeks (Wang et al., 2013).

• Group III (Empagliflozin group):

12 rats were fed HFD for 8 weeks then during the next 8 weeks of the study the rats received HFD plus empagliflozin suspension, at a daily dose of 10 mg/ kg (1.6 ml) by gastric tube (**Jojima et al., 2016**).

• Group IV (Silymarin group):

12 rats were fed HFD for 8 weeks then during the next 8 weeks of the study the rats received HFD plus silymarin suspension, at a daily dose of 100 mg/kg (1.4 ml) by gastric tube (**Huang et al., 2010**).

***** Outcome measures:

1- Biochemical studies:

(A) Collection of blood samples:

Blood samples were collected from the retroorbital venous plexus of rat eye by using heparinized capillary tubes. The collected blood was then centrifuged at 3000 round/minute for 30 minutes. Then the serum was transferred into clean vials and stored at -18° C for biochemical parameters determination and the abdomens of the rats were dissected and the livers were excised to measure the following parameters:

(B) Biochemical measurements:

Serum parameters:

- Alanine aminotransferase (ALT).
- Aspartate aminotransferase (AST).
- Triglycerides (TG).
- Cholesterol.

 \triangleright

- Random blood glucose.
- Serum insulin level.

Liver homogenate parameters:

Liver samples were homogenized for measurement of:

- Hepatic levels of malondialdehyde (MDA).
- Hepatic levels of reduced glutathione (GSH).
- Liver tissue content of triglycerides.
- Transforming growth factor β (TGF- β).

2- Body weight recordings:

Body weight was recorded to the rats of all groups at the beginning and at the end of the study.

3- Histopathology:

A small piece of liver was immediately fixed in 10% neutral buffered formalin then embedded in paraffin wax. For light microscopic analysis of liver histology, the paraffin-embedded liver tissues were cut into 4μ m sections, and standard hematoxylin-eosin (H & E) staining was performed. Hepatic fibrosis was assessed by Masson trichrome stain.

The liver sections were scored according to the NAFLD Activity Score (NAS), which includes the features of active liver injury.

The score is defined as the sum of the scores for steatosis (0-3), lobular inflammation (0-3), and ballooning (0-2); thus ranging from 0 to 8 (**Kleiner et al., 2005**).

Ten light microscopic fields were viewed on each section and scored for the severity of hepatic steatosis, inflammation, ballooning and fibrosis according to the following criteria:

> For hepatic steatosis:

Score 0: no fat.

Score 1: steatosis occupying less than 33% of the hepatic parenchyma.

Score 2: 34–66% of the hepatic parenchyma.

Score 3: more than 66% of the hepatic parenchyma.

> For inflammatory cell infiltration:

Score 0: none.

- Score 1: 1-2 foci/field.
- Score 2: 3-4 foci/field.

Score 3: more than 4 foci/field.

Hepatocellular ballooning

Score 1: few ballooned cells.

Score 2: marked ballooning.

The staging of **hepatic fibrosis** was investigated by Masson staining as:

Stage 1: Perisinusoidal / pericellular fibrosis; focally or extensively present (lesion in the central vein area).

Stage 2: Zone 3 perisinusoidal/ pericellular fibrosis with focal or extensive periportal fibrosis (lesion in the central vein area and expansion to the surrounding area).

Stage 3: Portal fibrosis with focal or extensive bridging fibrosis.

Stage 4: Cirrhosis (Brunt et al., 1999; Kleiner et al, 2005).

3. Results

The effects of treatment of NAFLD rats with empagliflozin (10 mg/kg, p.o. /day) and silymarin (100 mg/kg, p.o. /day) together with high fat diet (HFD) for 8 weeks preceded by 8 weeks during which the rats were receiving HFD only on Serum ALT, AST, total cholesterol, serum TG and random blood glucose levels: **High fat diet (group II)** significantly increased serum levels of ALT by (490.15 %), AST by (533.69 %), total cholesterol by (103.08 %), serum TG by (95.49 %) and random blood glucose by (157.49%) compared to normal control group (group I).

Empagliflozin 10 mg/ kg (group III) administration significantly decreased serum levels of ALT by (54.77 %), AST by (29.83 %), total cholesterol by (38.11 %), serum TG by (32.51 %) and random blood glucose by (33.61%) compared to HFD group (group II).

Silymarin 100 mg/ kg (group IV) administration significantly decreased serum levels of ALT by (61.06 %), AST by (36.81 %), total cholesterol by (21.61 %), serum TG by (12.46 %) and random blood glucose by (6.7 %) compared to HFD group (group II).



Figure (1): The effects of different studied groups on serum alanine aminotransferase (ALT) (U/ml):



Figure (2): The effects of different studied groups on serum aspartate amino transferase (AST) (U/ml):



Figure (3): The effects of different studied groups on total serum cholesterol (mg/ dl):



Figure (4): The effects of different studied groups on serum triglycerides (TG) (mg/ dl):



Figure (5): The effects of different studied groups on random blood glucose (mg/ dl):

a: Significantly different from negative control group.

b: Significantly different from HFD group.

d: Significantly different from HFD+ empagliflozin 10 mg/kg / day treated group.

g: Significantly different from HFD+ silymarin 100 mg/kg / day treated group.

2- The effects of treatment of NAFLD rats with empagliflozin (10 mg/kg, p.o. /day), and silymarin (100 mg/kg, p.o. /day) together with high fat diet HFD for 8 weeks preceded by 8 weeks during which the rats were receiving HFD only on hepatic level of MDA, GSH, TG and TGF- β :

High fat diet (group II) administration significantly increased hepatic levels of MDA by



Figure (6): The effects of different studied groups on hepatic malondialdhyde (MDA) (mmol/ gm tissue):



Figure (7): The effects of different studied groups on hepatic reduced glutathione (GSH) (mmol/ gm tissue):

(103.43 %), decreased GSH by (45.3 %), increased hepatic TG by (231.1 %) and TGF- β by (601.68 %) compared to normal control group (group I).

Empagliflozin 10 mg/ kg (group III) administration significantly decreased hepatic levels of MDA by (42.42 %), increased GSH by (32.01 %), deceased TG by (52.77 %) and TGF- β by (30.28 %) compared to HFD group (group II).

Silymarin 100 mg/ kg (group IV) administration significantly decreased hepatic levels of MDA by (44.60 %), increased GSH by (41.58 %), deceased TG by (41.95 %) and TGF- β by (41.3 %) compared to HFD group (group II).



Figure (8): The effects of different studied groups on hepatic triglycerides (TG) (mg/ gm tissue):



Figure (9): The effects of different studied groups on hepatic transforming growth factor- β (TGF- β) (mg/ dl): a: Significantly different from negative control group. b: Significantly different from HFD group.

d: Significantly different from HFD+ empagliflozin 10 mg/kg / day treated group.

g: Significantly different from HFD+ silymarin 100 mg/kg / day treated group.

The effects of treatment of NAFLD rats with empagliflozin (10 mg/kg, p.o. /day) and silymarin (100 mg/kg, p.o. /day) together with high fat diet HFD for 8 weeks preceded by 8 weeks of HFD only on body weight, Serum insulin level and NASH Score.

High fat diet (group II) significantly increased body weight by (18.14 %), serum insulin level by (156.72 %) and NASH Score by (377.64 %) compared to normal control group (group I).

Empagliflozin 10 mg/ kg (group III) administration significantly decreased body weight by (9.78 %), serum insulin level by (30.35 %) and NASH Score by (43.59 %) compared to HFD group (group II).

Silymarin 100 mg/ kg (group IV) administration non-significantly decreased body weight by (1.41 %), significantly decreased serum insulin level by (19.68 %) and NASH Score by (14.16 %) compared to HFD group (group II).







Figure (11): The effects of different studied groups on serum insulin (pg/ ml):



Figure (12): The effects of different studied groups on NAS score:

a: Significantly different from negative control group.

b: Significantly different from HFD group.

d: Significantly different from HFD+ empagliflozin 10 mg/kg / day treated group.

g: Significantly different from HFD+ silymarin 100 mg/kg / day treated group.

Histopathological findings

Group I (Negative control group): the pathological study of this group confirmed the clinical serological parameters. Liver section of this group showed normal structure and architecture (Haematoxylin & Eosin $\times 100$).

Group II (High fat diet group): liver section of HFD fed group II showed severe degree of micro and macrovescicular steatosis and severe hepatocellular ballooning with frequent foci of inflammatory cells (Haematoxylin & Eosin ×200). Liver section of HFD fed group II showed marked portal fibrosis (masson trichom stain×100).

Group III (Empagliflozin treated group): liver sections of rats treated with empagliflozin showed less frequent inflammatory foci and moderate steatosis (H & E x100). Liver sections of rats treated with empagliflozin showed mild perisinasoidal fibrosis (masson stain x100).

Group IV (Silymarin treated group): liver sections of rats treated with silymarin (group IX) showed marked steatosis and hydrobic changes together with inflammatory foci (H & E x100). Liver sections of rats treated with silymarin showed marked perisinasoidal fibrosis (masson stain x100).



Figure (13): Liver section of control group I showing normal structure and architecture (Haematoxylin & $Eosin \times 100$).





Figure (14-A): liver section of HFD fed group II showing severe degree of micro and macrovescicular steatosis and severe hepatocellular ballooning with frequent foci of inflammatory cells (black arrow) (Haematoxylin & Eosin ×200). **Figure (14-B):** Liver section of HFD fed group III showing marked portal fibrosis (yellow arrow) (masson trichom stain×100).



15-A)



Figure (15-A): Liver sections of rats treated with empagliflozin showing less frequent inflammatory foci and moderate steatosis (red arrows) (H & E x100). **Figure (15-B):** Liver sections of rats treated with empagliflozin showing mild perisinasoidal fibrosis (yellow arrow) (masson stain x100).





16-A)

Figure (16-A): Liver sections of rats treated with silymarin (group IX) showing marked steatosis and hydrobic changes together with inflammatory foci (black arrows) (H & E x100). **Figure (16-B):** Liver sections of rats treated with silymarin showing marked perisinasoidal fibrosis (yellow arrow) (masson stain x100).

4. Discussion

Non-alcoholic fatty liver disease (NAFLD) is a spectrum of liver disorders. It is a condition defined by the presence of steatosis in more than 5 % of hepatocytes (**Sanyal et al., 2011**) with a little or no alcohol consumption. NAFLD consists of the benign non-alcoholic fatty liver (NAFL), and the more severe non-alcoholic steato-hepatitis (NASH). NASH is a more progressive form of NAFLD and is characterized by steatosis, hepatocellular ballooning, lobular inflammation and almost always fibrosis (**Kleiner and Makhlouf, 2016**). In an effort to regenerate new cells,

The biological mechanism underlying steatosis occurrence and progression to NAFLD is not yet fully understood. Association of NAFLD with obesity, type 2 diabetes and metabolic syndrome suggests that; hyperglycemia, hyperlipidaemia, and insulin resistance are triggering factors in pathogenesis of NAFLD (Mookhan et al., 2014).

NAFLD can be considered as a hepatic manifestation of a metabolic syndrome. These findings have created a surge toward development of strategies to: control obesity, improve glycemic control, enhance insulin sensitivity and improve β -cell and hepatocyte function. Drug therapy has typically been focused on the management of associated risk factors such as diabetes, obesity, and hyperlipidemia, being predisposing factors for development of NAFLD (Mookhan et al., 2014).

Although a multiple animal models of NAFLD exist, high fat diet (HFD) administration has been a

NASH progresses to cirrhosis (Cobbina and Akhlaghi, 2017).

NAFLD prevalence is high, being stated in around 20 to 30% of the general population in studies based on imaging methods. For histological studies, in selected groups of patients with risk factors for this disease, the prevalence may be higher, with steatosis found in 70% of obese patients and 35% of non-obese individuals, while NASH is seen in 18.5% of obese and 3% of non-obese patients. In patients with type 2 diabetes mellitus (DM2), the frequency of fatty liver disease can reach 75% (**Binobaid et al., 2018**).

widely used model and was chosen in the present study (Asai et al., 2014).

In the present study, HFD significantly increased serum levels of ALT, AST, total serum cholesterol, serum TG, random blood glucose, hepatic levels of MDA, decreased hepatic levels of GSH, increased hepatic TGs, TGF- β , serum insulin and NASH score compared to normal control group. These results agree with Lin et al., 2011 and Yao et al., 2017.

Sodium-glucose co-transporter-2 (SGLT2) inhibitors are a new class of oral antidiabetic drugs, which reduce hyperglycemia by promoting urinary glucose excretion independently of the secretion or action of insulin (**Tahrani et al., 2013**). Empagliflozin is a highly selective SGLT2 inhibitor that improves glycemic control in patients with T2DM (**Grempler et al., 2012**). In a preclinical model of obesity, empagliflozin triggered weight loss associated with reduction of body fat that was probably secondary to calorie wasting in the urine (**Vickers et al., 2014**). Silymarin, extracted from the milk thistle, is a mixture of the three isomeric compounds silybin, silidianin, and silichristin (Feher et al., 2008). Some randomized controlled trials have shown beneficial effects of silymarin such as antioxidant therapy and improved survival against a variety of liver ailments (Polyak et al., 2010).

In the present study, administration of empagliflozin at a dose of 10 mg/ kg (group IV) ameliorated HFD induced NAFLD by significantly decreasing insulin resistance (decreased random blood glucose and serum insulin level), decreasing markers of acute liver injury (decreased serum ALT and AST) and decreasing serum and hepatic lipid content (total serum cholesterol, serumTG, hepatic TG and body weigh) and decreasing NAS score compared to HFD group (group II).

The results of the current study coincides with Jojima et al., 2016, whom studied the effects of empagliflozin 10 mg/ kg on steato-hepatitis in a novel model of non-alcoholic steato-hepatitis mouse and diabetes found that; empagliflozin significantly decreased serum serum levels of ALT, total cholesterol, TG, NAS score and body weight. They demonstrated that empagliflozin prevented the development of NASH in this model by its antiinflammatory and anti-fibrotic effects. Expression of mRNA for FAS and ACC1, genes involved in fatty acid production (lipogenesis) were significantly lower in empagliflozin treated group than the vechile group. Empagliflozin decreased hepatic fibrosis by decreasing collagen deposition significantly compared to the vehicle group. Empagliflozin reduced hepatic inflammation, which was confirmed by decreased expression of TNF-a mRNAs significantly in the empagliflozin group compared with the vehicle group.

Another study agrees with these results done by Xu et al., 2019, whom documented that, after 8 weeks on a high-fat diet (HFD), mice exhibited obesity, accompanied by insulin resistance and low-grade chronic inflammation. Treatment with empagliflozin 0.01% (w/w) significantly decreased serum serum levels of ALT, AST, total cholesterol, TG, fasting glucose, insulin, NAS score and body weight. They explained their results by that, treatment with empagliflozin increased energy expenditure (i.e: empagliflozin-treated mice exhibited higher oxygen consumption and exhaled more CO2 than the HFD-fed mice did, suggesting that treatment with empagliflozin enhances use of carbohydrates as well as fats under HFD conditions) suggested an increase in adaptive thermogenesis. Empagliflozin increased adipose tissue browning (the mRNA expression of brown fatselective genes was also significantly increased in the empagliflozin treated group). empagliflozin alleviated obesity-associated insulin resistance (as detected by GTT, ITT and serum insulin level) and inflammation (as detected by significant decrease in inflammatory and proinflammatory markers) in the WAT and liver of obese mice.

Empagliflozin consistently reduced liver TG content and plasma levels of TG and TC in HFD-fed mice, and these findings were associated with the suppression of lipogenic gene expression and upregulation of mitochondrial fatty acid β -oxidation genes. Moreover, empagliflozin tended to upregulate liver fibroblast growth factor-21 (FGF21) protein level and markedly increased levels of plasma ketone bodies and glycerol, suggesting that lipolysis is enhanced in empagliflozin-treated mice. In addition, empagliflozin significantly decreased levels of plasma ALT and AST (empagliflozin alleviated HFD-induced liver damage) **(Xu et al., 2019).**

Theese results also are in agreement with Adingupu et al., 2019, whom studied the effects of empagliflozin 1.5 mg/kg body weight/day on coronary microvascular function and cardiac contractility in ob/ob⁻ prediabetic mice and showed that. empagliflozin treated animals experienced а significant reduction in serum levels of ALT, AST, total cholesterol, TG, fasting glucose, insulin NAS score and body weight There was a statistical difference in liver steatosis score between treated and untreated mice. Empagliflozin reduced glycemia, promoted caloric loss via glucose excretion and reduced body weight gain. SGLT2 inhibitors mediate a metabolic switch from glucose to lipid utilization (Ferrannini et al., 2014).

This study also is in line with **Mizuno et al., 2018,** whom studied the effects of empagliflozin 10 mg/kg per day on the size and number of mitochondria after myocardial infarction in diabetic hearts and found that, empagliflozin significantly decreased serum glucose, total cholesterol, and triglyceride. The treatment with empagliflozin increased lipolysis.

But **Mizuno et al., 2018,** showed that, empagliflozin increased lipid droplets and TG level in the myocardium. **Mizuno et al., 2018,** explained that by, since the rate of fatty acid uptake into cardiomyocytes is dependent on the blood concentration of FFA, it is possible that the lipid accumulation induced by empagliflozin resulted from elevation of the circulating FFA level, possibly caused by the 24- h fasting induced lipolysis in the setting of their experiments. They speculated that reduced transport of FFA to the mitochondria might also be involved in the accumulation of lipid droplets in cardiomyocytes.

These results are in accordance with **Sawada et al.**, , as they studied the effects of empagliflozin on plasma triglyceride in diabetic patients with coronary artery disease and found that; after the 6 month

treatment, body weight and body fat percentage decreased significantly, plasma triglyceride, HbA1C level and fasting and postprandial plasma glucose levels were significantly decreased by treatment. Postprandial insulin secretion was also significantly suppressed and insulin resistance index was significantly decreased. Furthermore, fasting and postprandial triglyceride levels decreased significantly.

Sawada et al., 2019, noted that 6 months of treatment with empagliflozin improved glycemic obesity, insulin resistance. control. hypertriglyceridemia. In addition, it ameliorated inflammation. Improvement in both fasting and postprandial hypertriglyceridemia was the strongest independent factor predicting improvement of endothelial dysfunction in diabetic patients with CAD. treated with empaglifolozin. Empagliflozin indirectly improve insulin sensitivity by reducing glucose toxicity (Abdul-Ghani et al., 2011). Furthermore, weight reduction results from calorie loss because of glucosuria and lipid breakdown (Cai et al., 2017).

The results of the current study showed that, administration of silymarin at a dose of 100 mg/ kg ameliorated HFD induced NAFLD by significantly decreasing insulin resistance (decreased serum insulin level), decreasing markers of acute liver injury (decreased serum ALT and AST) and decreasing serum and hepatic lipid content (total serum cholesterol, serumTG and hepatic TG) and decreasing NAS score compared to HFD group (group II). Silymarin produced a non-significant change in random blood glucose and body weight as compared to HFD group.

This study is in accordance with that obtained by **Mahmoud et al., 2018,** whom studied the effects of silymarin 100 mg/kg/ day by means of oral gavage on high-fat-induced fatty liver in rats and found that, silymarin significantly decreased ALT, AST, total serum cholesterol, serum TG, hepatic TG, serum insulin level and NAS score as compared to HFD.

Multiple biochemical, metabolic and signal transduction pathways contribute to insulin resistance. It was found that protein glycation may play a role in insulin resistance by a variety of mecha-nisms, including generation of tumor necrosis factor-alpha, direct modification of the insulin molecule leading to its impaired action, generation of oxidative stress and impairment of mitochondrial function (Song et al., 2012). So, the beneficial effect of silymarin is mostly due improvement of insulin resistance (Mahmoud et al., 2018).

These results accordnates with a previous report, in which silymarin treatment was associated with a reduction of insulin resistance and a significant decrease in fasting insulin levels, suggesting an improvement of the activity of endogenous and exogenous insulin (Cacciapuoti et al., 2013), also Federico et al., 2006, showed that a new silybin vitamin E complex improves insulin resistance and liver damage in patients with NAFLD.

The results of the current work are in line eith **Kheiripour et al., 2019**. They studied the hepatoprotective effects of silymarin on liver injury and found that, silymarin resulted in a significant reduction of serum AST, ALT, random blood glucose, serum insulin and body weight.

Silymarin has many beneficial capabilities such as antioxidant, anti-inflammatory, immunemodulatory, and liver-regenerating capacities (**Surai** et al., 2015; Kheiripour et al., 2019).

Silymarin inhibits liver injury by maintaining the integrity of the plasma membrane, inhibits the secretion of liver enzymes in blood, and suppresses apoptosis in hepatocytes (Vargas-Mendoza et al., 2014; Kheiripour et al., 2019)

But this study doesn't agrees with that obtained by Ni and Wang, 2016, whom studied the effects of Silymarin on hepatic steatosis through regulation of lipid metabolism and oxidative stress in a mouse model of non-alcoholic fatty liver disease (NAFLD) and found that, silymarin produced a slight nonsignificant decrease in ALT, AST, body weight. But this study was conducted on mice that were fed with HFD for 20 weeks and treated with silymarin 30 mg/ kg only for 4 weeks only. But they showed that silvmarin significantly decreased serum total cholesterol. serum TG, hepatic TG and histopathological changes induced by HFD.

Gobalakrishnan et al., 2016, studied the effects of silybin on lipid profile in hypercholesterolaemic rats and found that treament with Silybin significantly decreased serum total cholesterol serum TG and hepatic TG.

In silybin treated rats the fecal excretion of bile acids was increased. Enhanced excretion of faecal bile acids by Silybin depletes the pool of bile acids, and hepatic bile acid synthesis increases. As a result, hepatic cholesterol declines, stimulating the production of hepatic LDL receptors leading to enhanced clearance of plasma LDL, IDL and indirectly that of VLDL. This could be a possible mechanism of hypolipidemic effect of Silybin (Gobalakrishnan et al., 2016).

In the present study, empagliflozin significantly ameliorated antioxidant and anti-inflammatory effects induced by NAFLD (as it decreased hepatic levels of MDA, TGF- β and significantly increased hepatic GSH level).

The results of this work are in accordance with that obtained by **Abdelhamid et al., 2020**, whom studied the effects of Empagliflozin on ethanolinduced liver injury and found that; empagliflozin significantly decreased hepatic level of MDA and increased hepatic GSH level. empagliflozin significantly decreased serum levels of alanine aminotransferase, aspartate aminotransferase, and alkaline phosphatase as markers of liver injury. Moreover, empagliflozin inhibited the release of proinflammatory cytokines, including TNF- α , IL-1 β , and IL-6 as markers of inflammation, via the downregulation of NF-kB. These changes were associated with an improvement in histopathological deterioration. In conclusion Abdelhamid et al., 2020 showed that, empagliflozin modulated alcohol-induced liver damage by suppressing oxidative stress and inflammatory responses.

These results suggest that empagliflozin exhibits remarkable free radical scavenging activities and can reverse the reduction in the activities of antioxidant enzymes. In accordance with the present results, previous studies have demonstrated that empagliflozin could act as an antioxidant by decreasing oxidative stress and enhancing antioxidative enzymes, independently from its known glucose-lowering activity (**Kabel et al., 2020 and Lahnwong et al., 2018**).

Empagliflozin showed many benefits potentially mediated through its ability to reduce inflammation, oxidative stress, apoptosis, mitochondrial dysfunction and ionic dyshomeostasis (Amin et al., 2020 and Yaribeygi et al., 2019).

The current study also coincides with Li et al., 2019, whom studied the effect of empagliflozin on myocardial oxidative stress and fibrosis in diabetic mice heart and found empagliflozin to decrease hepatic level of MDA and increase hepatic GSH level. They confirmed that, empagliflozin could effectively control blood glucose and reduce insulin release. In addition to its role in good glycemic control, empagliflozin also significantly lowers blood cholesterol and TG levels in diabetic mice.

Empagliflozin inhibited the classical TGF- β /Smad pathway and decreased cardiac oxidative stress (Li et al., 2019).

Regarding the antioxidant and antiinflammatory effects of silymarin, it significantly decreased hepatic level of MDA, TGF- β and increased hepatic GSH level.

In concidance with our results, **Mahmoud et al., 2018,** found that, administration of silymarin resulted in a significant reduction of hepatic MDA, TGF- β and increased hepatic GSH level significantly. Silymarin has demonstrated antioxidant properties, through stimulation of polymerase and RNA transcription, by protecting the cell membrane from radical-induced damage and blocking the uptake of toxins (Wellington et al., 2001).

This study is in line with **Yao et al., 2013**. They studied the effect and the probable mechanisms of silibinin in regulating insulin resistance in the liver of rats with non-alcoholic fatty liver and found that, compared with the model group; silybin treatment significantly decreased hepatic malondialdehyde (MDA) and increased hepatic GSH level, which coincides with the recent study. Also silibinin significantly decreased visceral fat, serum total cholesterol, TG, ALT, AST and IR.

This study also is in accordance with **Mahli et al., 2015,** whom studied the hepatoprotective effect of oral application of a silymarin extract in carbon tetrachloride-induced hepatotoxicity in rats and found that, silymarin resulted in a significant reduction of hepatic TGF- β compared to NAFLD non-treated group. They found that the silymarin extract had direct inhibitory effects on pro-inflammatory and pro-fibrogenic gene expression in HSCs in vitro. This indicates that direct effects on HSC also contribute to the in vivo hepatoprotective effects of silymarin, and further promote its potential as anti-fibrogenic agent also in chronic liver disease.

Histopathological study of empagliflozin treated group showed less frequent inflammatory foci, moderate steatosis and mild perisinasoidal fibrosis compared to HFD group.

This histopathology coincides with **Jojima et al., 2016** and **Xu et al 2017** whom found that; empagliflozin significantly improved fatty degeneration, inflammatory cell infiltration and hepatocellular ballooning and TG content..

Histopathological examination of silymarin treated group showed marked steatosis. hydrobic changes, inflammatory foci with marked perisinasoidal fibrosis.

This study agrees with **Mahmoud et al., 2018**, whom found that, administration of silymarin resulted in a significant decrease of pathological changes induced by high fat diet feeding and improvement of histopathological scores in liver samples showing mild to moderate steatosis and less inflammation and ballooning.

Silymarin has an anti-inflammatory effect by acting through different mechanisms as its antioxidant action, membrane stabilizing effect and inhibition of production or release of inflammatory mediators (Mahmoud et al., 2018).

Administration of silibinin for 6 weeks significantly improved fatty degeneration and inflammation (manifesting as less cytological ballooning, relatively regularly arranged cell cords and decreased lobular inflammation). In addition silymarin reduced fat accumulation in hepatocytes (Yao et al., 2013). This work also accords with that obtained by **Ni and Wang, 2016,** whom found that, silymarin significantly reversed HFD induced histopathologic changes that induced by HFD. silymarin could attenuate hepatic steatosis in a mouse model of NAFLD through regulation of lipid metabolism and oxidative stress, and benefit to the circulation system

As compared to empagliflozin, in the present study, silymarin decreased ALT, AST, and hepatic MDA more significantly, but vildagliptin significantly decreased total serum cholesterol, serum TG, random blood glucose and hepatic TG more significantly compared to silymarin. Silymarin increased GSH and decreased TGF- β more significantly as compared to empagliflozin. Empagliflozin dcreased body weight significantly while silymarin produced non-significant decrease in the body weight. Empagliflozin reduced serum insulin level and NASH score more significantly than silymarin.

The beneficial effects of empapagliflozin could be explained by that, empagliflozin is a potent oral anti-diabetic agent that inhibits sodium-glucose cotransporter 2 (SGLT-2). By inhibiting this transporter, SGLT-2 inhibitors promote urinary glucose excretion, which, in turn, decreases blood levels of glucose and improves insulin resistance in patients with type 2 diabetes leading to down-regulation of carbohydrate-responsive element-binding protein (ChREBP), a transcription factor responsible for activating the machinery for fatty acid synthesis. Improvement in insulin resistance (hyperinsulinemia) results in down-regulation of SREBP-1c and the blockage of hepatic lipogenesis (Kuchav et al., 2018).

Empagliflozin markedly decreased obesity induced inflammation in the liver of diet induced obese (DIO) mice (**Xu et al., 2017**). These findings suggest that SGLT2 inhibitors, particularly empagliflozin, improve insulin resistance partially by attenuating chronic inflammation in obese and diabetic subjects (**Xu and Ota, 2018**).

Consistent with other SGLT2 inhibitors, administering empagliflozin to HFD-induced obese mice mitigates weight gain and fatty liver. The underlying mechanism for the weight reduction depends partially on increased energy expenditure and enhanced fatty acid oxidation. Empagliflozin increases oxygen consumption and tends to elevate carbon dioxide exhalation, leading to increased sugar and fat utilization (**Xu et al., 2017**).

Empagliflozin increases the plasma LDL-C level concomitant with higher FFAs and total ketone body levels, which suggests that inhibiting SGLT2 induces ketogenesis and a metabolic switch toward lipid oxidation to counterbalance the carbohydrate restriction (**Briand et al., 2016**).

Administering empagliflozin increases fatty acid oxidation by altering the expression of adiponectin and leptin in epididymal WAT. The adipose tissuespecific adipokines; leptin and adiponectin are involved in the regulation of food intake and energy homeostasis (Adya et al., 2015).

Plasma leptin stimulates fatty acid esterification to triglycerides and causes an even greater increase in hydrolysis so that there is a net efflux of fatty acids from the cells. By contrast, adiponectin exerts its insulin-sensitizing effects by increasing βoxidation of fatty acids and reducing serum triglyceride and FFA levels, thus indirectly improving insulin sensitivity. Furthermore, leptin and adiponectin interact with AMPK, which regulates fatty acid and energy metabolism. Administering empagliflozin increases adiponectin mRNA expression and down-regulates leptin expression in epididymal WAT and contributes to fat lipolysis and energy expenditure. Taken together, these findings indicate that empagliflozin improves abnormal lipid metabolism and obesity by enhancing fat and sugar utilization and increasing fatty acid oxidation (Xu and Ota, 2018).

Zhao et al., 2016, showed elevation of adiponectin down-regulates the renal SGLT2 by activating PPAR δ , which in turn reduces reabsorption of sodium and glucose (**Zhao et al., 2016**). On the other hand, SGLT2 inhibitors increase the expression of adiponectin in diabetic subjects and obese model (**Xu et al., 2017; Tanizawa et al., 2013**). Empagliflozin promotes adiponectin expression in WAT (**Xu et al., 2017**).

Treatment with SGLT2i(s) results in decreases in both glucose and insulin levels (especially in those with T2DM) which lead to a large decrease in hepatic de novo lipid synthesis (**Daniele et al., 2016**).

Glucagon-secreting alpha cells of pancreatic islets express SGLT2, and inhibition of the SGLT2 cotransporter (which presumably reduces intracellular glucose concentration in these cells an important signal for glucagon release) results in increased secretion and blood levels of glucagon (**Daniele et al., 2016; Wang et al., 2018).**The high glucagon levels (and elevated glucagon/insulin ratio) lead to stimulation of β -oxidation (and hepatic ketone production and elevated blood ketone levels) and cause a shift from carbohydrate to fatty acid metabolism and help reduce liver triglyceride content ((**Daniele et al., 2016**).

In keeping with the importance of glucagon in the pathogenesis of NAFLD, a recent study showed that while a reduction in glucagon receptor expression (using antisense technology) in patients with T2DM improved glycemic control, it also increased their ALT levels and significantly increased hepatic fat content (Morgan et al., 2019). As compared to vildagliptin treated group, empagliflozin treated group showed non-significant changes in serum ALT, AST, significant decrease in total serum cholesterol, serum TG, non-significant changes in random blood glucose, hepatic MDA, GSH, significant decrease in hepatic TG, non-significant changes in hepatic TGF- β , body weight, significant decrease in serum insulin level and non-significant decrease in NASH score.

Silybin could be an insulin sensitizer; as it is able to reduce intrahepatic fat accumulation, lobular inflammation, ballooning and serum fat, as well as to improve homeostasis model assessment-IR index (HOMA-IR) and insulin tolerance test (ITT). Moreover, silybin has an important role in reducing visceral fat accumulation, in inducing lipolysis through the transcription of the adipose triglyceride lipase (ATGL) gene and inhibiting gluconeogenesis for silencing of some genes involved in the fat metabolic pathway (Yao et al., 2013).

Silybin has low bioavilability if administered per orally. It reduces the absorption of fat contained in HFD mediated by the formation of non-absorbable complexes, rather than depending on its real role in interrupting the pathogenetic mechanisms that are responsible for NAFLD (**Christodoulou et al., 2015**).

The treatment with increasing doses of silybin (16, 40 and 100 μ g/mL), in vitro, is able to generate an increase in glucose captation, induced by insulin in a model of palmitate-induced insulin-resistance on myoblast C2C12 cells, in which the role of silybin is crucial (Li et al., 2015).

Conclusion:

Empagliflozin and silymarin had ameliorated HFD induced biochemical, inflammatory, oxidative stress and histopathological changes, but empagliflozin was more effective and more significant. So, more research is needed to investigate the effectiveness of these drugs in NAFLD.

References

- Abdelhamid A, Elsheakh AR, Abdelaziz R and Abdul-Ghani MA, Norton L and Defrozo RA (2011): Role of sodium-glucose cotransporter type 2 (SGLT2) inhibitors for the treatment of type 2 diabetes. Endocr Rev. 2011;32:515-531.
- Abul-Fadlel KA, Mohammed NA, Al-Sayed RM, Abdul-Rahman M and Farag AI (2020): Effect of spexin treatment on cardiometabolic changes in obese type 2 diabetic rats E. Al-Azhar Med. J.Vol. 49(2), April, 2020, 735-758.
- 3. Adingupu D, Göpel S, Grönros J, Behrendt M, Sotak M, Miliotis T, Dahlqvist U, Ming Gan L & Rylander A (2019): SGLT2 inhibition with empagliflozin improves coronary microvascular

function and cardiac contractility in prediabetic ob/ob^{-/-} mice. Cardiovascular Diabetology volume 18, Article number: 16 (2019).

- 4. Aghahoseini F, Alihemmati A, Hosseini L & Badalzadeh R (2020): Vildagliptin ameliorates renal injury in type 2 diabetic rats by suppressing oxidative stress. Journal of Diabetes & Metabolic Disorders.
- 5. Bedogni G, Bellentani S, Miglioli L et al (2006): The fatty liver index: a simple and accurate predictor of hepatic steatosis in the general population. BMC Gastroenterol. 6:33.
- Briand F, Mayoux E, Brousseau E, Burr N, Urbain I, Costard C, Mark M and Sulpice T (2016): Empagliflozin, via switching metabolism toward lipid utilization, moderately increases LDL cholesterol levels through reduced LDL catabolism. Diabetes; 65 (7):2032–8.
- Brunt EM, Janney CG, Di Bisceglie AM, Neuschwander-Tetri BA & Bacon BR (1999): Nonalcoholic steatohepatitis: a proposal for grading and staging the histological lesions. The American journal of gastroenterology; 94(9), 2467-2474.
- Cai X, Ji L, Chen Y, Yang W, Zhou L, Han X, et al (2017): Comparisons of weight changes between sodium-glucose cotranspoter 2 inhibitors treatment and glucagon-like peptide-1 analogs treatment in type 2 diabetes patients: a meta-analysis. J Diabetes Investig. 2017;8:510-517.
- Chang M-w, Chen C-h and Chen Y-c, et al (2015): Sitagliptin protects rat kidneys from acute ischemia-reperfusion injury via upregulation of GLP-1 and GLP-1 receptors. *Acta Pharmacol Sin*. 2015;36:119–130.
- Chitapanarux T, Tienboon P, Suwalee P and Donrawee L (2009): Open-labeled pilot study of cysteine-rich whey protein isolate supplementation for nonalcoholic steatohepatitis patients. J Gastroenterol Hepatol. 2009;24:1045– 1050.
- Christodoulou E, Kechagia I.A, Tzimas S, Balafas E, Kostomitsopoulos N, Archontaki H, Dokoumetzidis A and Valsami G (2105): Serum and tissue pharmacokinetics of silibinin after per os and i.v. administration to mice as a HP-β-CD lyophilized product. Int. J. Pharm; 493, 366–373.
- 12. Cusi K (2016): Treatment of patients with type 2 diabetes and non-alcoholic fatty liver disease: current approaches and future directions. Diabetologia; 59 (6):1112–1120.
- 13. Daniele G, Xiong J, Solis-Herrera C, Merovci A, Eldor R, Tripathy D, DeFronzo RA, Norton L and Abdul-Ghani M (2016): Dapagliflozin enhances fat oxidation and ketone production in patients

with type 2 diabetes. Diabetes Care; 39(11):2036–2041.

- 14. Daniele G, Xiong J, Solis-Herrera C, Merovci A, Eldor R, Tripathy D, DeFronzo RA, Norton L and Abdul-Ghani M (2016): Dapagliflozin enhances fat oxidation and ketone production in patients with type 2 diabetes. Diabetes Care; 39(11):2036– 2041.
- Davis L.M., Pei Z., Trush M.A., Cheskin L.J., Controreggic C., Mccullougk K., et al (2006): Bromoc-riptine reduces steatosis in obese rodent models. J. Hepa-tol., 45: 439-44, 2006.
- 16. De Luca C and Olefsky JM (2008): Inflammation and insulin resis-tance. *FEBS Lett* 582:97–105.
- 17. Duez H, Cariou B and Staels B (2012): DPP-4 inhibitors in the treatment of type 2 diabetes. Biochem Pharmacol; 83(7): 823–832.
- El-Far M, Negm A, Wahdan M (2015): Promising Biopharmaceutical Use Of Silymarin And Silibinin As Antidiabetic Natural Agents In Streptozotocin-Induced Diabetic Rats: First Comparative Assessment. World Journal Of Pharmacy And Pharmaceutical Sciences. 4(2):07-28.
- 19. El-Lakkany NM, Salah el-Din SH, Sabra ANAA, Hammam OA & Ebeid FAL (2016): Coadministration of metformin and N-acetylcysteine with dietary control improves the biochemical and histological manifestations in rats with.
- El-Marasy SA, Abdel-Rahman RF and Abd-Elsalam RM (2018): Neuroprotective effect of vildagliptin against cerebral ischemia in rats. Naunyn-Schmiedeberg's. Arch Pharmacol. 2018;391:1133–1145.
- 21. Fan K, Wu K, Lin L, Ge P, Dai J, He X, Hu K and Zhang L (2107): Metformin mitigates carbon tetrachloride-induced TGF- β 1/Smad3 signaling and liver fibrosis in mice. Biomedicine & Pharmacotherapy. Volume 90, June 2017, Pages 421-426.
- 22. Federico A, Trappoliere M, Tuccillo C, de Sio I, Di Leva A, Del Vecchio Blanco C and Loguercio C (2006): A new silybin-vitamin E-phospholipid complex improves insulin resistance and liver damage in patients with non-alcoholic fatty liver disease: Preliminary observations; 55, 901–902.
- 23. Feher J and Lengyel G (2008): Silymarin in the treatment of chronic liver diseases: past and future]. Orv Hetil; 149: 24132418.
- Ferrannini E, Baldi S, Frascerra S, Astiarraga B, Heise T, Bizzotto R, Mari A, Pieber TR, Muscelli E (2016): Shift to fatty substrate utilization in response to sodium-glucose votransporter 2 inhibition in subjects without diabetes and patients with type 2 diabetes. Diabetes; 65(5):1190–1195.

- 25. Ferrannini E, Muscelli E, Frascerra S, Baldi S, Mari A, Heise T, Broedl UC and Woerle HJ (2014): Metabolic response to sodium-glucose cotransporter 2 inhibition in type 2 diabetic patients. J Clin Invest. 2014;124(2):499–508.
- 26. Field FJ, Born E, Murthy S and Mathur SN (2001): Gene expression of sterol regulatory element-binding proteins in hamster small intestine. J. Lipid Res; 42, 1–8.
- 27. Filhoulaud G, Guilmeau S, Dentin R, Girard J and Postic C (2013): Novel insights into ChREBP regulation and function. Trends Endocrinol Metab; 24(5):257–268.
- 28. Girard J, Perdereau D, Foufelle F, Prip-Buus C and Ferré P (1994): Regulation of lipogenic enzyme gene expression by nutrients and hormones. FASEB J; 8:36–42.
- 29. Gobalakrishnan s, Asirvatham ss and Janarthanam v (2016): Effect of Silybin on Lipid Profile in Hypercholesterolaemic Rats. J Clin Diagn Res. 2016 Apr; 10(4): FF01–FF05.
- Guo J, Zhou Y, Cheng Y, Fang W, Hu G, Wei J, Lin Y, Man Y, Guo L, Sun M, Cui Q and Li J (2018): Metformin-Induced Changes of the Coding Transcriptome and Non-Coding RNAs in the Livers of Non-Alcoholic Fatty Liver Disease Mice. Cell Physiol Biochem 2018;45:1487–1505.
- Gutierrez-Repiso C, Rodriguez-Pacheco F, Garcia-Arnes J, Valdes S, Gonzalo M, Soriguer F, Moreno-Ruiz FJ, Rodriguez-Cañete A, Gallego-Perales JL and Alcain-Martinez G (2015): The expression of genes involved in jejunal lipogenesis and lipoprotein synthesis is altered in morbidly obese subjects with insulin resistance. Lab. Investig; 95, 1409–1417.
- 32. Hawley SA, Ford RJ, Smith BK, Gowans GJ, Mancini SJ, Pitt RD, Day EA, Salt IP, Steinberg GR and Hardie DG (2016): The Na+/glucose cotransporter inhibitor canagliflozin activates AMPK by inhibiting mitochondrial function and increasing cellular AMP levels. Diabetes; 65(9):2784–94.
- 33. Horst KWT and Serlie MJ (2017): Fructose consumption, lipogenesis, andnon-alcoholic fatty liver disease. Nutrients; 9 (9):981.
- 34. Huang B., Ban X., He J., Tong J., Tian J. and Wang Y (2010): Hepatoprotective and antioxidant activity of ethanolic extracts of edible lotus (Nelumbo nucifera Gaertn.) leaves. Food Chemistry; 120 (3): 873- 878, 2010.
- 35. Hussain M, Babar MZM, Hussain MS, and Lubna Akhtar (2016): Vildagliptin ameliorates biochemical, metabolic and fatty changes associated with non-alcoholic fatty liver disease. Pak J Med Sci; 32(6): 1396–1401.

- 36. Huypens P, Quartier E, Pipeleers D and Van De Casteele M (2005): Metformin reduces adiponectin protein expression and release in 3T3-L1 adipocytes involving activation of AMP activated protein kinase. *Europ J Pharmacol.* 2005;518:90–95.
- 37. Irace C, Casciaro F, Scavelli FB, Oliverio R, Cutruzzola A, Cortese C, et al. (2018): Empagliflozin influences blood viscosity and wall shear stress in subjects with type 2 diabetes mellitus compared with incretin-based therapy. Cardiovasc Diabetol. 2018;17(1):52.
- 38. Jalali M, Rahimlou M, Mahmoodi M, Moosavian SP, Symonds M, kJalali R, Zare M, HadiImanieh M and Stasi C (2020): The effects of metformin administration on liver enzymes and body composition in non-diabetic patients with nonalcoholic fatty liver disease and/or non-alcoholic steatohepatitis: An up-to date systematic review and meta-analysis of randomized controlled trials. Pharmacological Research. Volume 159, September 2020, 104799.
- Jayanta Paul (2020): Recent advances in noninvasive diagnosis and medical management of non-alcoholic fatty liver disease in adult. Egyptian Liver Journal, volume 10, Article number: 37 (2020)
- 40. Junker AE, Gluud L, Holst JJ, Knop FK and Vilsbøll T (2016): Diabetic and nondiabetic patients with nonalcoholic fatty liver disease have an impaired incretin effect and fasting hyperglucagonaemia. JIM; 279(5):485–493.
- 41. Kabel AM, Estfanous RS and Alrobaian MM (2020): Targeting oxidative stress, proinflammatory cytokines, apoptosis and toll like receptor 4 by empagliflozin to ameliorate bleomycin-induced lung fibrosis. Respiratory physiology & neurobiology. 2020;273:103316.
- 42. Kamal SM (2014): Anti-Oxidant and Anti-Inflammatory Effects of Vildagliptin in NonAlcoholic Fatty Liver Disease of Mice. Kamal. Int J Med Nano Res; 1:1. ISSN: 2378-3664.
- 43. Kawano Y and Cohen DE (2013) Mechanisms of hepatic triglyceride accumulation in nonalcoholic fatty liver disease. *J Gastroenterol.* 48: 434-441.
- 44. Kazemi R, Aduli M, Sotoudeh M, Malekzadeh R, Seddighi N, Sepanlou SG and Merat S (2102): Metformin in Nonalcoholic Steatohepatitis: A Randomized Controlled Trial. Middle East J Dig Dis. 2012 Jan; 4(1): 16–22.
- 45. Khalil R, Shata A, Abd El-Kader E, Sharaf H, Abdo WS Amin NA and Saber S (2020): Vildagliptin, a DPP-4 inhibitor, attenuates carbon tetrachloride-induced liver fibrosis by targeting

ERK1/2, p38 α , and NF- κ B signaling. Toxicology and Applied Pharmacology. Volume 407, 15 November 2020, 115246.

- 46. Kheiripour N, Karimi J, Khodadadi I, Tavilan H, Goodarzi MT and Hashemnia M (2019): Hepatoprotective Effects of Silymarin on Liver Injury via Irisin Upregulation and Oxidative Stress Reduction in Rats with Type 2 Diabetes. Iran J Med Sci. 2019 Mar; 44(2): 108–117.
- Khowailed EA, Seddiek HA, Mahmoud MM, Rashed L, Ibrahim F (2018): Effect of metformin on Sirtuin-1 disorders associated with diabetes in male rats. Alexandria Journal of Medicine. Volume 54, Issue 4, December 2018, Pages 373-381.
- 48. Kita Y, Takamura T, Misu H, Ota T, Kurita S, Takeshita Y, Uno M, Matsuzawa-Nagata N, Kato K and Ando H (2012): Metformin prevents and reverses inflammation in a non-diabetic mouse model of nonalcoholic steatohepatitis. PLoS ONE 7; e43056.
- 49. Kleiner DE and Makhlouf HR (2016): Histology of Nonalcoholic Fatty Liver Disease and Nonalcoholic Steatohepatitis in Adults and Children. Clin Liver Dis; 20:293–312.
- 50. Kleiner DE, Brunt EM, Van Natta M, Behling C, Contos MJ, Cummings OW & Yeh M (2005): Design and validation of a histological scoring system for nonalcoholic fatty liver disease. Hepatology; 41(6), 1313-1321.
- 51. Kotronen A, Peltonen M, Hakkarainen A et al (2009): Prediction of non-alcoholic fatty liver disease and liver fat using metabolic and genetic factors. Gastroenterology 137:865–872.
- 52. Kuchay MS, Krishan S, Mishra SK, Farooqui KJ, Singh MK, Wasir JS, Bansal B, Kaur P, Jevalikar G, Gill HK, Choudhary NS and Mithal A (2018): Effect of Empagliflozin on liver fat in patients with type 2 diabetes and nonalcoholic fatty liver disease: a randomized controlled trial (E-LIFT Trial). Diabetes Care; 41(8):1801–1808.
- 53. Kumashiro N, Erion DM, Zhang D, Kahn M, Beddow SA, Chu X, Still CD, Gerhard GS, Han X, Dziura J, Petersen KF, Samuel VT and Shulman GI (2011): Cellular mechanism of insulin resistance in nonalcoholic fatty liver disease. Proc Natl Acad Sci USA; 108(39):16381–16385.
- Lahnwong S, Chattipakorn SC and Chattipakorn N (2018): Potential mechanisms responsible for cardioprotective effects of sodium–glucose cotransporter 2 inhibitors. Cardiovascular diabetology. 2018;17:101.
- 55. Lee SS, Park SH, Kim HJ et al (2010): Noninvasive assessment of hepatic steatosis:

prospective comparison of the accuracy of imaging examinations. J Hepatol 52:579–585.

- 56. Li C, Zhang J, Xue M, Li X, Han F, Liu X, Xu L, Lu Y, Cheng Y, Li T, Yu X, Sun & Chen L (2019): SGLT2 inhibition with empagliflozin attenuates myocardial oxidative stress and fibrosis in diabetic mice heart. Cardiovascular Diabetology volume 18, Article number: 15.
- 57. Li CL, Zhao LJ, Zhou XL, Wu HX and Zhao JJ (2015): Effect of glucagon like peptide-1 receptor agonists and dipeptidyl peptidase-4 inhibitors for the treatment of non-alcoholic fatty liver disease. *J Huazhong Univ Sci Technol Med* Sci. 2015;35(3):333–336.
- 58. Loomba R, Lutchman G, Kleiner DE, Ricks M, Feld J, Borg B, Modi A, Nagabhyru P, Sumner AE, Liang TJ, and Hoofnagle JH (2009): Clinical trial: pilot study of metformin for the treatment of non-alcoholic steatohepatitis. Aliment Pharmacol Ther. 2009 Jan; 29(2): 172–182.
- Macauley M, Hollingsworth KG, Smith FE, Thelwall PE, Al-Mrabeh A, Schweizer A, Foley JE & Taylor R (2015): Effect of vildagliptin on hepatic steatosis. J. Clin. Endocrinol. Metab; 100, 1578–1585.
- Mahli A, Koch A, Czech B, Peterburs P, Lechner A, Haunschild J, Müller M & Hellerbrand C (2015): Hepatoprotective effect of oral application of a silymarin extract in carbon tetrachloride-induced hepatotoxicity in rats. Clinical Phytoscience volume 1, Article number: 5.
- 61. Mahmoud H, Fatma A, Mahmoud F and Hashim G (2018): The Beneficial Effects of Silymarin in Treatment of Experimentally Induced Non-Alcoholic Fatty Liver Disease in Rats. Med. J. Cairo Univ; Vol. 86, No. 2, March: 1055-1064.
- 62. Marchetti P, Del Guerra S, Marselli L, Lupi R, Masini M, Pollera M, et al (2004): Pancreatic islets from type 2 diabetic patients have functional defects and increased apoptosis that are ameliorated by metformin. *J Clin Endocrinol Metab.* 2004;89(11):5535–5541.
- 63. Marin V, Gazzin S, Gambaro SE, Ben M, Calligaris S, Anese M, Raseni A, Avellini C, Giraudi PJ, Tiribelli C and Rosso N (2017): Effects of Oral Administration of Silymarin in a Juvenile Murine Model of Non-alcoholic Steatohepatitis. Nutrients. 2017 Sep; 9(9): 1006.
- 64. Meng S, Yang F, Yueqiu Wang Y, Qin Y, Xian H, Che H and Wang L (2019): Silymarin ameliorates diabetic cardiomyopathy via inhibiting TGF-β1/Smad signaling. Cell Biol Int. 2019 Jan;43(1):65-72.
- 65. Michael H, Martin F and Standl E (2006): Vildagliptin-An oral dipeptidyl peptidase-4

inhibitor for type 2 diabetes. US Endocrine Dis; 2:75-81.

- 66. Miyagawa K, Kondo T, R. Goto R et al., (2013): "Effects of combination therapy with vildagliptin and valsartan in a mouse model of type 2 diabetes," *Cardiovascular Diabetology*, vol. 12, no. 1, p. 160, 2013.
- 67. Mizuno M, Kuno A, Yano T, Miki T, Oshima H, Sato T, Nakata K, Kimura, Y Tanno M, and Miura T (2018): Empagliflozin normalizes the size and number of mitochondria and prevents reduction in mitochondrial size after myocardial infarction in diabetic hearts. Physiol Rep; 6(12): e13741.
- Mookkan J, De S, Shetty P, Kulkarni NM, Devisingh V, Jaji MS, Lakshmi VP, Chaudhary S, Kulathingal J, Rajesh NB, and Narayanan S (2014): Combination of vildagliptin and rosiglitazone ameliorates nonalcoholic fatty liver disease in C57BL/6 mice. Indian J Pharmacol; 46(1):46-50.
- 69. Morgan ES, Tai L-J, Pham NC, Overman JK, Watts LM, Smith A, Jung SW, Gajdošík M, Krššák M, Krebs M, Geary RS, Baker BF, Bhanot S (2019): Antisense inhibition of glucagon receptor by IONIS-GCGR rx improves type 2 diabetes without increase in hepatic glycogen content in patients with type 2 diabetes on stable metformin therapy. Diabetes Care; 42:4.
- 70. Ni X and Wang H (2016): Silymarin attenuated hepatic steatosis through regulation of lipid metabolism and oxidative stress in a mouse model of non-alcoholic fatty liver disease (NAFLD). Am J Transl Res 8(2):1073-1081.
- 71. Ni X and Wang H (2016): Silymarin attenuated hepatic steatosis through regulation of lipid metabolism and oxidative stress in a mouse model of non-alcoholic fatty liver disease (NAFLD). Am J Transl Res 8(2):1073-1081.
- 72. Petta S, Gastaldelli A, Rebelos E, Bugianesi E, Messa P, Miele L, Svegliati-Baroni G, Valenti L and Bonino F (2016): Pathology of non-alcoholic fatty liver disease. Int J Mol Sci; 17(12):2082.
- Polyak SJ, Morishima C, Lohmann V, Pal S, Lee DY, Liu Y, Graf TN and Oberlies NH (2010): Identification of hepatoprotective flavonolignans from silymarin. Proc Natl Acad Sci U S A; 107: 5995-5999.
- 74. Postic C, Girard J (2008): Contribution of de novo fatty acid synthesis to hepatic steatosis and insulin resistance: lessons from genetically engineered mice. J Clin Invest; 118(3): 829-838.
- 75. Poynard T, Ratziu V, Naveau S et al (2005): The diagnostic value of biomarkers (SteatoTest) for the prediction of liver steatosis. Comp Hepatol. 4:10.

- 76. Refaat R, Sakr A, Salama M, and El Sarha A (2016): Combination of Vildagliptin and Pioglitazone in Experimental Type 2 Diabetes in Male Rats. DRUG DEVELOPMENT RESEARCH. 77: 300–309 (2016)
- 77. Reichetzeder C, Websky K and Tsuprykov O, et al (2017): Head-to-head comparison of structurally unrelated dipeptidyl peptidase 4 inhibitors in the setting of renal ischemia reperfusion injury. *Br J Pharmacol.* 2017; 174:2273–2286.
- 78. Rizzo MR, Marfella R, Barbieri M and Paolisso G (2012): Reduction of oxidative stress and inflammation by blunting daily acute glucose fluctuations in patients with type 2 diabetes. *Diabetes Care*. 2012; 35:2076–2082.
- 79. Rösen P and Wiernsperger NF (2006): Metformin delays the manifestation of diabetes and vascular dysfunction in Goto-Kakizaki rats by reduction of mitochondrial oxidative stress. *Diabetes Metab Res Rev.* 2006; 22(4):323–330.
- Salman ZK, Refaat R, Selima E, El Sarha A and Ismail MA (2013): The combined effect of metformin and L-cysteine on inflam-mation, oxidative stress and insulin resistance in streptozotocin-induced type 2 diabetes in rats. *Eur J Pharmacol*. 714:448–455.
- Sanyal AJ, Brunt EM, Kleiner DE, Kowdley KV, Chalasani N, Lavine JE, Ratziu V and Mccullough A (2011): Endpoints and clinical trial design for nonalcoholic steatohepatitis. Hepatology; 54: 344–53.
- 82. Sawada T, Uzu K, Hashimoto N, Onishi T, Takaya T, Shimane A, Taniguchi Y, Yasaka Y, Ohara T and Kawai H (2019): Empagliflozin Ameliorating Effect of on Plasma Triglyceride: An Association with Endothelial Function Recovery in Diabetic Patients with Coronary Artery Disease. J Diabetes Metab; Vol. 10 Iss. 5 No: 826.
- Sherif IO, Als-haalan AA and Al-Shaalan NH (2020): Renoprotective effect of vildagliptin following hepatic ischemia/reperfusion injury. Ren Fail. 2020; 42(1): 208–215.
- 84. Shirakawa J, Fujii H, Ohnuma K, Sato K, Ito Y and Kaji M (2011): Diet induced adipose tissue inflammation and liver steatosis are prevented by DPP-4 inhibition in diabetic mice. *Diabetes*. 2011;60(4):1246–1257.
- Silverman JF, O'Brien KF, Long S, Leggett N, Khazanie PG, Pories WJ, Norris HT, Caro JF (1990): Liver pathology in morbidly obese patients with and without diabetes. Am J Gastroenterol; 85(10):1349–1355.

- 86. Song F. and Schmidt A.M (2012): Glycation and Insulin Resistance. Arteriosclerosis, thrombosis, and vascular biology, 32 (8): 1760-1765, 2012.
- Suddek GM (2020): Empagliflozin ameliorates ethanol-induced liver injury by modulating NFκB/Nrf-2/PPAR-γ interplay in mice. Life Sciences, Volume 256, 1 September 2020, 117908.
- Surai PF (2015): Silymarin as a Natural Antioxidant: An Overview of the Current Evidence and Perspectives. Antioxidants (Basel) 2015; 4:204–47.
- Sviklāne L, Olmane E, Dzērve Z et al (2018): Fatty liver index and hepatic steatosis index for prediction of non-alcoholic fatty liver disease in type 1 diabetes. J Gastroenterol Hepatol 33:270– 276.
- 90. Thong-Ngam D, Samuhasaneeto S, Kulaputana O and Klaikeaw N (2007) N-acetylcysteine attenuates oxidative stress and liver pathology in rats with non-alcoholic steatohepatitis. *World J Gastroenterol*. 13: 5127-5132.
- 91. Toda N and Nakanishi-Toda M (2007): Nitric oxide: ocular blood flow,glaucoma, and diabetic retinopathy. *Prog Retinal Eye Res.* 26:205–238.
- 92. Van Stee MF, deGraaf AA and Groen AK (2018): Actions of metformin and statins on lipid and glucose metabolism and possible benefit of combination therapy. Cardiovasc. Diabetol; 2018, 17, 94.
- 93. Vargas-Mendoza N, Madrigal-Santillan E, Morales-Gonzalez A, Esquivel-Soto J, Esquivel-Chirino C, Garcia-Luna YG-RM, et al (2014): Hepatoprotective effect of silymarin. World J Hepatol. 2014;6:144–9.
- 94. Verma S, Jensen D, Hart J et al (2013): Predictive value of ALT levels for non-alcoholic steatohepatitis (NASH) and advanced fibrosis in nonalcoholic fatty liver disease (NAFLD). Liver Int 33:1398–1405.
- 95. Wang D, Luo Y, Wang X, Orlicky DJ, Myakala K, Yang P and Levi M (2018): The sodiumglucose cotransporter 2 inhibitor dapagliflozin prevents renal and liver disease in western diet induced obesity mice. Int J Mol Sci;19(1):137.
- 96. Wang W, Zhao C, Zhou J, Zhen Z, Wang Y & Shen C (2013): Simvastatin ameliorates liver fibrosis via mediating nitric oxide synthase in rats with non-alcoholic steatohepatitis-related liver fibrosis. PLoS One; 8(10), e76538.
- 97. Wellington K. and Jarvis B. Silymarin: A review of its clinical properties in the management of hepatic disorders. Bio. Drugs, 15: 465-489. [Pub. Med.], 2001.
- 98. Wenhua Du, Xiaoqing Ma, Shanshan Shao, Chunxiao Yu, Lifang Zhao1, Li Fang, Lingyan

Zhou and Fei Jing (2019): Vildagliptin reduces lipid levels possibly by mechanisms associated with HMGCR and CYP7A1. Int J Clin Exp Med 2019;12(9):11657-11665.

- 99. Wierzbicki AS and Oben J (2012): Nonalcoholic fatty liver disease and lipids. Curr Opin Lipidol. 2012;23(4):345–52.
- 100. Xiaoqing Ma, Wenhua Du, Shanshan Shao, Chunxiao Yu, Lingyan Zhou and Fei Jing (2018): Vildagliptin Can Alleviate Endoplasmic Reticulum Stress in the Liver Induced by a High Fat Diet. Biomed Res Int. 2018; 2018: 5045182.
- 101. Xu L & Ota T (2018): Emerging roles of SGLT2 inhibitors in obesity and insulin resistance: Focus on fat browning and macrophage polarization. ADIPOCYTE; VOL. 7, NO. 2, 121–128.
- 102. Xu L, Nagata N, Chen G, Nagashimada M, Fen Zhuge F, Ni Y, Sakai Y, Kaneko S and Ota T (2019): Empagliflozin reverses obesity and insulin resistance through fat browning and alternative macrophage activation in mice fed a high-fat diet. BMJ Open Diab Res Care; 7:e 000783.
- 103. Xu L, Nagata N, Nagashimada M, Zhuge F, Ni Y, Chen G, Mayoux E, Kaneko S and Ota T (2017): SGLT2 inhibition by empagliflozin promotes fat utilization and browning and attenuates inflammation and insulin resistance by polarizing M2 macrophages in dietinduced obese mice. E Bio Medicine; 20:137–149.
- 104. Yao J, Zhi M, Gao X, Hu P, Li C and Yang X (2013): Effect and the probable mechanisms of silibinin in regulating insulin resistance in the liver of rats with non-alcoholic fatty liver. Braz. J. Med. Biol. Res; 46, 270–277.
- 105. Yaribeygi H, Butler AE, Atkin SL, Katsiki N and Sahebkar A (2019): Sodium–glucose cotransporter 2 inhibitors and inflammation in

1/22/2021

chronic kidney disease: Possible molecular pathways. Journal of cellular physiology. 2019; 234:223-30.

- 106.Zahedi AS, Ghasemi A and Azizi F (2008): Serum nitric oxide metabo-lites in subjects with metabolic syndrome. *Clin Biochem.* 41:1342– 1347.
- 107. Zarghani S, Abbaszadeh S, Alizadeh M, Rameshrad M, Garjani A, Soraya H (2018: The Eeffect of Metformin Combined with Calcium-Vitamin D3Against Diet-Induced Nonalcoholic Fatty Liver Disease. Advanced Pharmaceutical Bulletin; 8(X), X-X.
- 108. Zhang Q, Xiao X, Zheng J, Li M, Yu M, Ping F, Wang T and Wang X (2018): A Possible Mechanism: Vildagliptin Prevents Aortic Dysfunction through Paraoxonase and Angiopoietin-Like 3. BioMed Research International.
- 109. Zhao Y, Gao P, Sun F, Li Q, Chen J, Yu H¹, Li L, Wei X, He H, Lu Z, Wei X, Wang B, Cui Y, Xiong S, Shang Q, Xu A, Huang Y, Liu D and Zhu Z (2016): Sodium intake regulates glucose homeostasis through the PPARdelta/adiponectin-mediated SGLT2 pathway. Cell Metab; 23 (4):699–711.
- 110. Zhou YJ, Zhou YF, Zheng JN et al (2017): NAFL screening score: a basic score identifying ultrasound-diagnosed non-alcoholic fatty liver. Clin Chim Acta 475:44–50.
- 111. Zhu X, Yan H, Xia M, Chang X, Xu X, Wang L, Sun X, Lu Y, Bian H, Li X, and Gao X (2018): Metformin attenuates triglyceride accumulation in HepG2 cells through decreasing stearylcoenzyme A desaturase 1 expression. Lipids Health Dis. 2018; 17: 114.